Who am I?

My name is Lorenzo Rovigatti and my email is lorenzo.rovigatti@uniroma1.it

How did I get her

- 2009 Laurea in Fisica here (w
- 2012 PhD in Materials Science
- 2013-2014 Post-doc here (with
- 2014-2016 Post-doc at the U
- 2016-2017 Post-doc at the U
- 2017-2018 Researcher (RTD)
- 2018-now Tenure-track resea

What are my research

- Soft matter (including some bid
- Self-assembly (colloids, protei)
- DNA nanotechnology
- Polymeric assemblies (star polymers, microgels)

vith FS)
ce here (with FS)
th FS)
niversity of Vienna
niversity of Oxford
A) at CNR
archer (RTDB) here
archer (RTDB) here nterests?
archer (RTDB) here Interests? ophysics)
archer (RTDB) here Interests? ophysics) ns, nucleic acids)
archer (RTDB) here interests? ophysics) ns, nucleic acids)

Some pictures

Self-assembly vs. phase separation



Patchy particles





All-DNA materials



Polymeric objects (microgels)

This is not a scientific talk at a conference, it is a lesson

Please do interrupt me to ask questions

SELF-ASSEMBLY AND PHASE SEPARATION IN THE CELL

OR All the arguments you studied in a single place![†]

[†]"all" may be an exaggeration, but you get the point

Lorenzo Rovigatti

Physics Department, Sapienza University of Rome

Invited lecture for the Soft and Biological Matter course, January 14th 2021

A physicist take on biology^T

It's a matter of organisation all the way down

Larger



[†]I apologise for this gross oversemplification



$pprox 10~{ m nm}$



McGuffee and Elcock, PLoS Comput. Biol. 2010

Organelles



Perform specialised tasks
Well-separated from the rest of the cell
Membrane-bound
Maintain their "identity" over time
Communicate through molecular signals

More organelles?

REPORT

Germline P Granules Are Liquid Droplets That Localize by Controlled Dissolution/Condensation

Clifford P. Brangwynne^{1,2,3}, Christian R. Eckmann¹, David S. Courson³, Agata Rybarska¹, Carsten Hoege¹, Jöbin Gharakhani²...



$Droplets \rightarrow phase \ separation$

Science 26 Jun 2009: Vol. 324, Issue 5935, pp. 1729-1732 DOI: 10.1126/science.1172046

What kind of phase separation?



A (gas-like) highly-diluted phase coexist with a (liquid-like) dense phase

Protein liquid-liquid phase separation (LLPS) = colloidal gas-liquid phase separation

Membraneless organelles



E. Gomes and J. Shorter, J. Mol. Bio. 2018

- Also known as biomolecular condensates
- Liquid-like (they can and do flow, cfr. Brangwynne *et al*)
- Made of multivalent (intrinsically-disordered) proteins and/or RNA
- Act as reservoirs of biomolecules or as microreactors
- The mechanisms behind their formation are linked to the pathogenesis of several diseases (*e.g.* Alzheimer's, ALS)

On-the-fly compartmentalisation

- Phase separation relies on a subtle interplay between entropy and enthalpy
- Slightly changing some conditions can suppress/enhance phase separation
- Condensates can quickly adapt to environmental changes!

A visual example

- We take a multi-component mixture \rightarrow we need to set many different interactions (red-blue, red-green, green-blue, etc.)
- For this particular choice the resulting system is homogeneous
- We change a single interaction \rightarrow phase separation



LLPS in proteins

- Abnormal LLPS is connected to many diseases Alzheimer's, Huntington's disease, ALS, frontotemporal dementia
- LLPS seems to be involved in transcriptional control
 - It helps controlling when, how and in what quantity proteins are synthesised
- Some types of condensates act as scaffolds that concentrate other molecules LLPS is used in the cell to generate "micro-reactors"
- Mechanical properties affect the morphology and localisation of the condensates Condensates are influenced by how stiff/flexible is the cell's local environment • Relative concentrations are connected to the overall biocondensate stability the thermodynamics can become really complicated

Some motivating problems

- The interior of the cell contains thousands of different macromolecules • Evolutionary pressure optimises mutual interactions in the proteome • In vivo and in vitro experiments don't always match (Jain and Vale, Nature 2017)

More motivation?

Letter to the Editor Published: 08 September 2020

Liquid–liquid phase separation by SARS-CoV-2 nucleocapsid protein and RNA

Hui Chen, Yang Cui, Xuling Han, Wei Hu, Min Sun, Yong Zhang, Pei-Hui Wang, Guangtao Song, Wei Chen

Cell Research 30, 1143–1145(2020) Cite this article

Article Open Access Published: 27 November 2020

Nucleocapsid protein of SARS-CoV-2 phase separates into RNA-rich polymerase-containing condensates

Adriana Savastano, Alain Ibáñez de Opakua, Marija Rankovic & Markus Zweckstetter 🖂

Nature Communications 11, Article number: 6041 (2020) Cite this article

Article | 4 December 2020 | 👌 OPEN ACCESS

C SOURCE DATA | TRANSPARENT PROCESS

SARS-CoV-2 nucleocapsid protein phase-separates with RNA and with human hnRNPs

Theodora Myrto Perdikari, Anastasia C Murthy, Veronica H Ryan, Scott Watters, Mandar T Naik, Nicolas L Fawzi 💿 🞽

Author Information EMBO J (2020) 39: e106478 | https://doi.org/10.15252/embj.2020106478

Where is the physics?

EVERYWHERE, AT EVERY LEVEL

- Everything is hydrated \rightarrow thermodynamics of water
- Proteins and nucleic acids are polymers \rightarrow polymer physics (elasticity, etc.)
- Proteins and nucleic acids bind reversibly ightarrow self-assembly, gels
- Cells are very crowded \rightarrow entropic effects (depletion, nematic transition, etc.) • The final boss: living organisms are, by definition, out of equilibrium

We need simple[†] systems and models we can play with to disentangle all these effects!

[†] in a reductionist sense

Our idea[†]

Convince a living organism to stochastically express artificial proteins designed to phase separate

The artificial proteins (expressed in a yeast strain through plasmids) should

- be extraneous to the cell

- bond through a lock-and-key (L&K) mechanism • have a tunable bonding strength (*i.e.* a tunable affinity) • be multivalent (*e.g.* can bind to multiple partners)

The goal: build a synthetic toolkit to study LLPS in vivo

[†] Emmanuel Levy's ingenious idea, to be really honest

The building blocks



- Two components (A and B) that can form up to 4 and 2 bonds, respectively
- Their size is chosen so that multiple bonding is unlikely (if not impossible)
- The L&K attraction strength can be tuned with point mutations

A closer look



Do they really form condensates?



Transmission & scanning electron micoscropy images

A preliminary check

Control sample: no phase separation if the L&K attraction is disabled



L&K enabled: the components co-localise and phase separate!





Quantifying the phase behaviour

We draw a *negative* of the low- ρ part of the phase diagram!



NB: the high-concentration part of the phase diagram cannot be accessed

Investigating the dynamics

FRAP: fluorescence recovery after photobleaching





A simple (but not too simple) model

The main ingredients

- No water \rightarrow implicit solvent, effective interactions
- No (or few) internal degrees of freedom \rightarrow spheres or quasi-rigid bodies
- Multivalency (a.k.a. limited valence in the colloidal field)
- A lock-and-key (L&K) mechanism (a.k.a. bond specificity)



Bianchi et al., Phys. Chem. Chem. Phys. 2017

Simple models \rightarrow complex behaviour

Limited valence





Simple models \rightarrow complex behaviour

Self-assembly vs. phase separation



F. Sciortino, A. Giacometti and G. Pastore, Phys. Rev. Lett. (2009)

J. Russo, J. M. Tavares, P. I. C. Teixeira, M. M. Telo da Gama and F. Sciortino, Phys. Rev. Lett. (2011)

L. Rovigatti, J. M. Tavares and F. Sciortino, Phys. Rev. Lett. (2013)

The theory

What you know: the F per particle of a self-assembling ideal gas of valence M

$$eta f\equiv rac{eta F}{N}=eta f_{
m id}+M\ln(1$$

It can be generalised to the case where the reference system is not an ideal gas

$$eta f = rac{eta F_{ ext{ref}}}{N} + rac{eta F_{ ext{bond}}}{N} = eta f_{ ext{ref}} + eta f_{ ext{bond}}$$

A few definitions when we deal with L&K binary mixtures

- x is the composition (fraction of A particles, for instance)
- M_A and M_B is the number of sites of species A and B
- X_A and X_B is the probability that a A or B site is unbonded
- B_2^{AA} , B_2^{AB} , B_2^{BB} are the second virial coefficients between the species

$$(-p_b)+rac{Mp_b}{2}$$

The theory

The two terms of the free energy now become

$$egin{aligned} eta f_{ ext{ref}} &= eta f_{ ext{id}} + x \ln(x) + (1-x) \ eta f_{ ext{bond}} &= x \left(M_A \ln(X_A) - rac{M_A}{2}
ight) \ &+ (1-x) \left(M_B \ln(X_B) - rac{M_A}{2}
ight) \end{aligned}$$

where βf_{ex} is the excess free energy (e.g. the hard-sphere excess free energy).

In the case of soft systems (ρ is the overall density)

$$eta f_{ ext{ex}} pprox
ho \left(x^2 B_2^{AA} + 2x(1-x)B_2^A
ight)$$

Free energy \rightarrow whole phase diagram



 $AB_{2}^{AB} + (1-x)^{2}B_{2}^{BB}$

Connecting theory and simulations

The second virial coefficients can be estimated as

$$B_{ij} = -rac{1}{2} \int_{0}^{\infty} 4 \pi r^2 \left(e^{-eta V_{ij}(r)} - 1
ight) dr$$

where $V_{ij}(r)$ is the reference interaction potential between species i and j

 X_A and X_B are connected to the bonding free energy Δ

$$\Delta = 4\pi \int_0^\infty g_{
m ref}(r) \langle f_{12}(r)_{\omega_1\omega_2}
angle r^2 dr$$

- $g_{\rm ref}(r)$ is the reference hard-sphere fluid pair correlation function
- $\langle f_{12}(r)_{\omega_1\omega_2} \rangle$ is the angle-averaged Mayer function at distance r

These quantities can be computed with two-body simulations

The coarse-graining strategy



Adding simulations

- Binary mixture of divalent and tetravalent particles, varying ho, x, T (\propto affinity $^{-1}$)
- Sedimentation simulations for the phase diagrams in and out of equilibrium
- Constant-volume simulations to compare to dynamic data
- Explore how changing the model affects the thermodynamics



cles, varying ho, x, T (\propto affinity⁻¹) and out of equilibrium namic data ermodynamics

Sedimentation

We fit the equation of state to extract the coexisting ho_2 and ho_4



Experimental phase diagrams

Affinity (interaction strength)



- The coexistence region is pprox symmetric with respect to stoichiometry conditions
- It enlarges as affinity (*i.e.* bond strength) increases
- Something happens at high affinities: out-of-equilibrium effects?

espect to stoichiometry conditions ases quilibrium effects?

Theoretical phase diagrams



- The coexistence region is symmetric with respect to stoichiometry conditions
- It enlarges as affinity (*i.e.* bond strength) increases

We reproduce all the equilibrium qualitative trends

pect to stoichiometry conditions eases

Numerical phase diagrams

Numerical phase diagrams in and out of equilibrium



We see the same qualitative shift measured in experiments

Out-of-equilibrium effects!

Affinity (interaction strength)



- FRAP data on single cells
- Large spread between the curves, especially at low affinity
- At large affinities there is \approx no recovery after 25 seconds



especially at low affinity very after 25 seconds

Model extensions

We can build on the model to assess the role

- of non-specific attractions
- of defects (example: some of tetramers can form only 3 bonds)
- of the flexibility/geometrical arrangements of the patches



ents of the patches

Conclusions

- We can engineer synthetic "playground" biomolecular condensates The phase behaviour of the system can be directly measured • Notwithstanding the complexity of the cell environment, experiments agree with coarse-grained simulations/theory, in and out of equilibrium

- The system can be used to test hypothesis and develop new methods (see e.g. Mc Laughin et al, Mol. Biol. Cell (2020))
- It's a starting point: we can add more ingredients!

Further questions?

- Ask me now!
- Contact me: lorenzo.rovigatti@uniroma1.it
- Read the paper: M. Heidenreich et al., Nat. Chem. Biol. 16, 939 (2020)

Some lines of research[†]

The role of polymer elasticity in the formation of condensates

- The presence of an elastic network alters the phase behaviour of liquids How does this relate to the formation/dissolution of molecular condensates? • Use simulations to test (and improve!) theories and compare to experiments

Interactions between DNA nanostructures

- DNA can be used to build nano-structures, machines and devices • We know how to predict and control the hybridisation between small strands What about large all-DNA nanostructures (DNA origami)?



[†]"Advertising"

Acknowledgements

Work done in collaboration with:

- E. Locatelli (University of Vienna)
- J. P. K. Doye (University of Oxford)
- S. K. Nandi (Tata Institute of Fundamental Research)



- M. Heidenreich (Weizmann Institute)
- S. A. Safran (Weizmann Institute)
- E. D. Levy (Weizmann Institute)

The building blocks









4B4K



D5



1VPX



Extending the model

Small patches \rightarrow single-bond-per-patch but slow equilibration



Big patches \rightarrow easier to find partners but multiple bonding possible





The best of both worlds

The FS[™] trick



Configuration



The value of λ controls the behaviour

- $\lambda \geq 1
 ightarrow ext{single-bond-per-patch}$
- $\lambda = 1 \rightarrow$ free swapping!

F. Sciortino, Eur. Phys. J. E (2017), L. Rovigatti et al., Macromolecules (2018)



 $-\epsilon$

 $\mathbf{0}$

- -2ϵ

Bond swapping \rightarrow no arrest

Systems can be equilibrated down to T o 0



L. Rovigatti et al., Macromolecules (2018)

Free lunch, for once

Faster equilibration for $\lambda = 1$



What is the influence of λ ?

Systems with $\lambda = 1$ and 10 have the same phase diagrams



Direct coexistence

The recipe

- 1. Equilibrate at small pressure (the specific value is not important) 2. Enlarge the box along one direction (here z) 3. Compute the density profile and extract the coexisting densities



Sedimentation vs. direct coexistence

$$x=rac{
ho_4}{
ho_2+
ho_4}$$
 , $ho=
ho$



Less numerically demanding but no out-of-equilibrium options

 $\rho_2 + \rho_4$

Equilibrium phase diagrams

The phase diagram enlarges as the bond strength (affinity) grows, as in the experiments



FRAP pictures



С

